

Optical Environment Sensing

-Multimodal Measurement of Cell Culture Conditions-



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培養面のpH・温度・Ca²⁺を長時間・安定計測!!

Background

Regeneration of bone *in vivo*

Bone mineralization *in vivo*

Critical defect (Ø9 mm) in Rat calvaria bone

Suzuki O et al, Biomaterials (2006), Suzuki O et al, Biomaterial (2009), Masuda T et al, Tissue Eng (2010)

Conventional simulator for catheter ablation

OCP conversion to HA

ALP activity

Measurement of **culture condition during conversion** is important.

Concept

Multimodal measurement of cell culture condition by fluorescence sensors

Measurement of pH and Ca²⁺ change during OCP conversion to HA

Environmental change during conversion of OCP to HA

(1) Hydrolysis
 $5Ca_8H_2(PO_4)_6 \cdot 5H_2O \rightarrow 4Ca_{10}(PO_4)_6(OH)_2 + 6H_3PO_4 + 17H_2O$

(2) [Ca²⁺] consumption
 $Ca_8H_2(PO_4)_6 \cdot 5H_2O + 2Ca^{2+} \rightarrow Ca_{10}(PO_4)_6(OH)_2 + 3H_2O + 4H^+$

OCP conversion to HA

Conventional method for physiological sensing

Method	Electrode	Fluorescence
Photo		
Multimodal sensing	Possible	Possible
Spatial resolution	Low	High

Fluorescence dyes for Multimodal sensing

	Dye	Excitation nm	Emission nm
pH	FITC	488	515
Ca ²⁺	Fura-2	360	500
Temp.	Rhodamine B	561	610

Fabrication of hydrogel sensor

Fabrication process

- (1) Dropping PEGDA solution
- (2) Contact of mask
- (3) Exposure UV
- (4) Development
- (5) Fabrication of other sens

Materials of sensors

- DI water : 55%
- PEGDA : 30%
- Omnirad 1173 : 5%
- Fura-2+DI water : 5%
- FITC+DMSO : 5%
- Rhodamine b : 1wt%

Fabricated fluorescence sensor

Experiment

Calibration of fluorescence sensor

Ca²⁺ calibration

pH calibration

Temp. calibration

pH and Ca²⁺ sensing by sensor

Cell culture on sensors

Cell: MC3TC-E1

Reference

T. Kobayashi, H. Maruyama, S. Omata, T. Masuda, F. Arai, Optical Environment Sensing -Multimodal Measurement of Cell Culture Conditions-, Robomech2018, 1A1-L04, 2018

Conclusions

Multimodal sensing of Ca²⁺, pH during OCP conversion was succeeded.

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